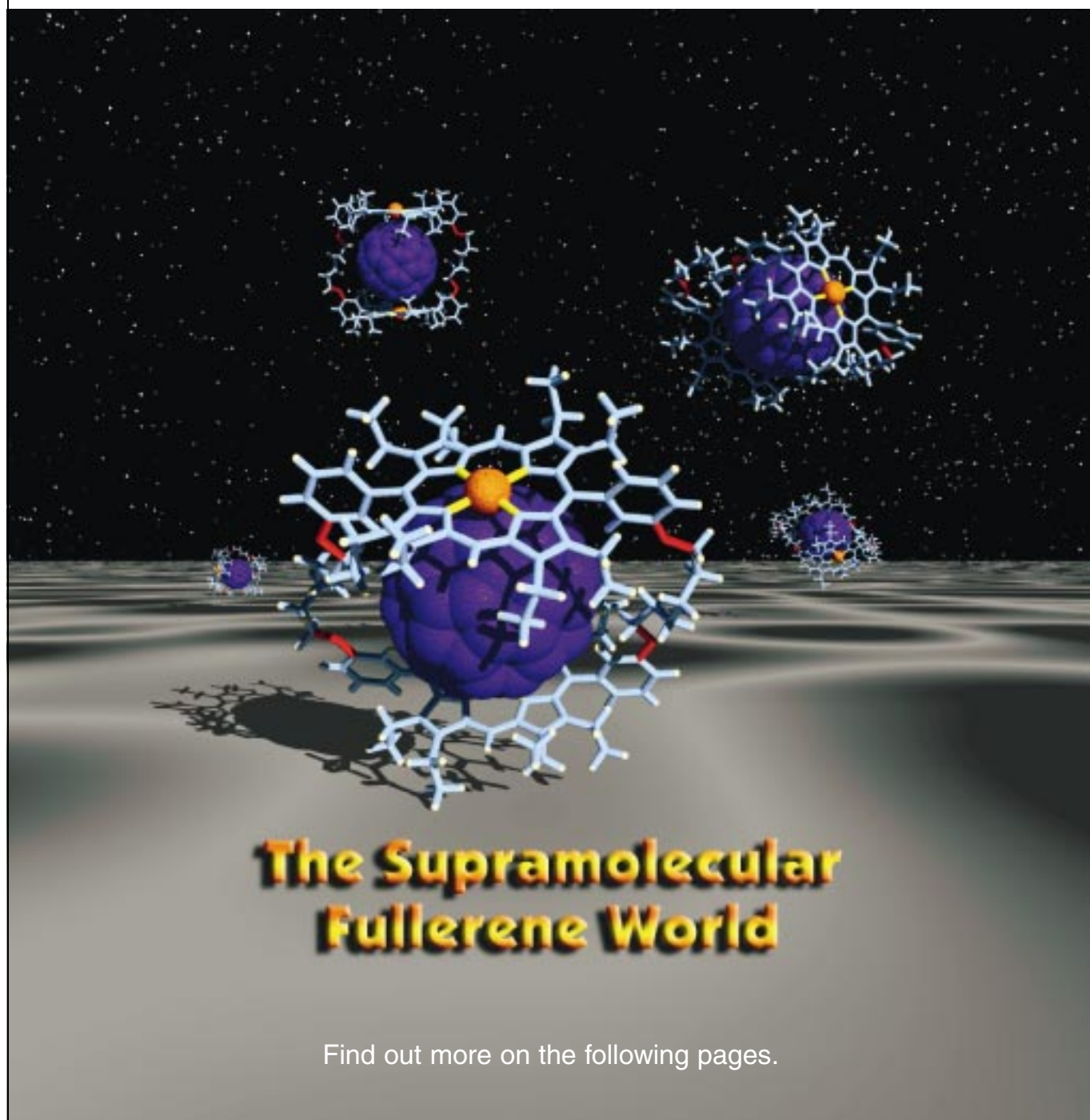


Supramolecular Architectonics with Carbon Nanoclusters

A host-guest interaction between fullerenes and cyclic dimers of metalloporphyrins results in the formation of doubly layered π -electronic supramolecules, whose thermodynamic stability and dynamics in guest exchange can be tuned by the selection of the metal ions at the outer porphyrin layer. In particular, the host molecule with rhodium ions exhibits an extremely high affinity toward fullerenes.



Cyclic Dimers of Metalloporphyrins as Tunable Hosts for Fullerenes: A Remarkable Effect of Rhodium(III)**

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With regard to multiwalled graphitic nanoparticles such as carbon nanopolyhedrons,^[1] nanocapsules,^[2] and buckyonions,^[3] wrapping of fullerenes in large π -conjugated molecular envelopes is an interesting subject, which may extend the π -electronic conjugation of fullerenes in a three-dimensional fashion. Recently, we found that a cyclic dimer of zinc porphyrin (**1-Zn**) forms an inclusion complex with C_{60} (**1-Zn** \supset C_{60}), of which the association constant K_{assoc} of $6.7 \times 10^5 \text{ M}^{-1}$ is the highest value reported to date in organic

media.^[4] This finding prompted us to investigate whether the central metal ions play an essential role in this supramolecular interaction.^[5] Here we report the results of a study on the interaction of fullerenes such as C_{60} and C_{70} with a series of metalloporphyrin cyclic dimers (**1-M**; $M = 2\text{H}$, Co, RhMe, Ni, Cu, Ag, and Zn), with emphasis on the extremely high affinity of the host molecule for Group 9 metal ions such as rhodium(III).^[6]

X-ray crystallography is informative of fullerene–metalloporphyrin interactions.^[7] By slow diffusion of acetonitrile into a CH_2Cl_2 solution of a mixture of C_{60} and **2-Zn**, a black prismatic crystal of a 1:1 complex of C_{60} with **2-Zn**, appropriate for X-ray crystallography, was obtained.^[8] Figure 1

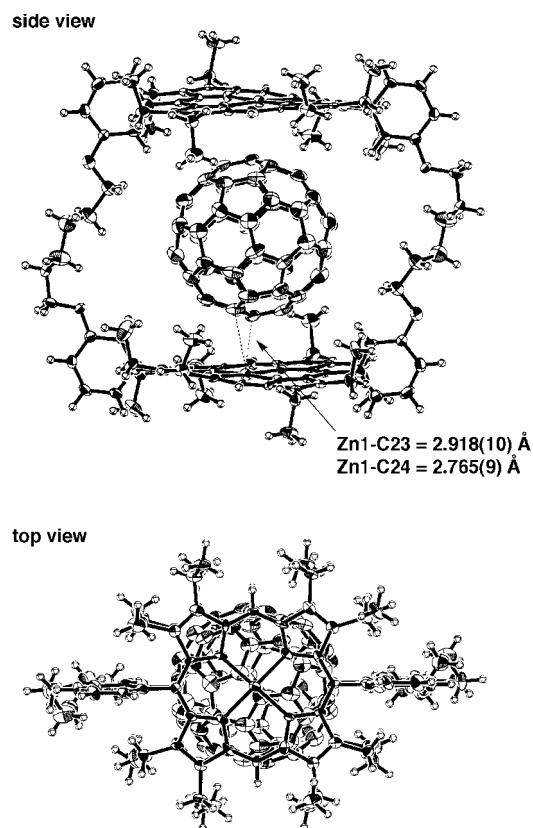
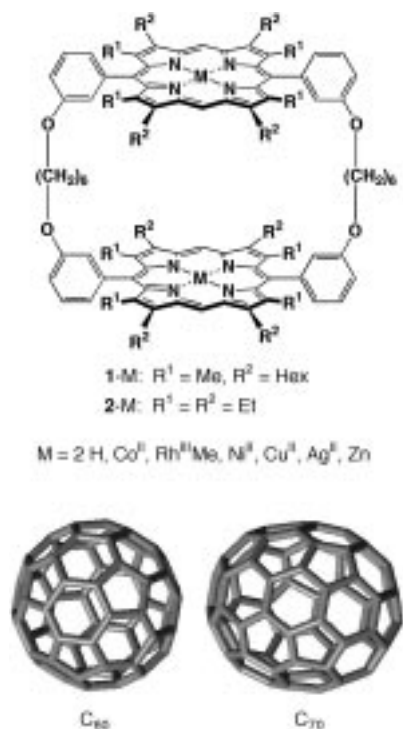


Figure 1. ORTEP plot of an inclusion complex of C_{60} with **2-Zn** (**2-Zn** \supset C_{60}).

shows an ORTEP plot of the complex at -110°C ,^[9] where **2-Zn** in the crystal adopts a C_i -symmetric conformation, the inversion center of which is shared with that of included C_{60} . The hexamethylene linkers of **2-Zn** are folded to adjust the porphyrin–porphyrin distance, and the porphyrin macrocycles are slightly distorted from planarity so as to accommodate the convex shape of C_{60} . One of the 5:6 ring-juncture C–C bonds of C_{60} is located on the central metal ion. The distances between the zinc atom and these two unique carbon atoms are 2.765(9) and 2.918(10) Å, which are clearly shorter than the sum of the van der Waals radii (3.09 Å).^[10] Furthermore, the distances between the pyrrole α -C atoms of the porphyrin moieties and three proximal carbon atoms of C_{60} (3.320–3.346 Å) are slightly but definitely shorter than the typical interlayer distance of graphite (3.354 Å).^[11] These observations suggest the presence of a π interaction between

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the zinc porphyrin moieties and the included fullerene in the solid state.^[7]

Similarly to our previous study,^[4] we conducted spectroscopic titration of the metalloporphyrin cyclic dimers bearing Group 9–11 metal ions (**1-M**; M = Co, Ni, Cu, Ag) in addition to free-base **1-2H**, with C₆₀ in benzene at 25 °C, in which the Soret absorption band of each host molecule showed a clear red shift with a decrease in absorbance.^[12] From the observed spectral change profiles, the association constants K_{assoc} were evaluated as shown in Figure 2a, whereby K_{assoc} of **1-2H** is

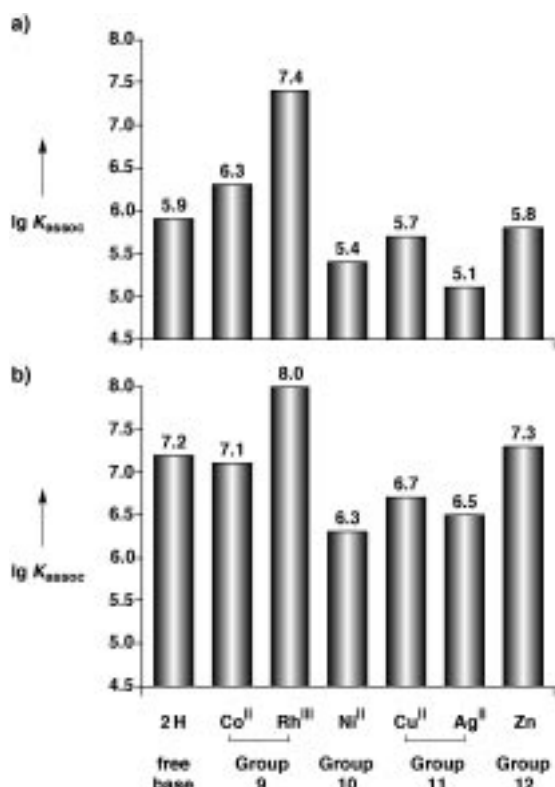


Figure 2. Association constants K_{assoc} of metalloporphyrin cyclic dimers **1-M** with C₆₀ (a) and C₇₀ (b) in benzene at 25 °C.

comparable to that of **1-Zn**,^[4] while **1-Ni**, **1-Cu**, and **1-Ag** with Group 10 and 11 metal ions are inferior to these two host molecules. On the other hand, a host molecule with Group 9 metal ions such as **1-Co** is clearly superior to all these host molecules. Therefore, we synthesized **1-RhMe** with rhodium(III) ions at the porphyrin center. Interestingly, the K_{assoc} value of **1-RhMe** with C₆₀ ($2.4 \times 10^7 \text{ M}^{-1}$) was even higher than that of **1-Co**.^[13] For the interaction with C₇₀, the affinities of the cyclic host molecules **1-M** displayed essentially the same dependency on the central metal ion as in Figure 2a, whereby the observed K_{assoc} values were nearly or more than one order of magnitude larger than those with C₆₀ (Figure 2b). In particular, **1-RhMe** again exhibited an extremely high affinity; the K_{assoc} value of about 10^8 M^{-1} apparently exceeds the experimental upper limit of accuracy for the spectroscopic titration. The K_{assoc} values of **1-2H**, **1-Co**, and **1-Zn** were also greater than 10^7 M^{-1} , whereas the other three host molecules with Group 10 and 11 metal ions (**1-Ni**, **1-Cu**, and **1-Ag**) were inferior to the above host molecules ($K_{\text{assoc}} < 10^7 \text{ M}^{-1}$). With

respect to the extremely high affinity of **1-RhMe** toward fullerenes, the inclusion complex **1-RhMe** ⊂ C₆₀ eluted as a single component not only in GPC on Polystyragel (CHCl₃) but also in HPLC on Buckyprep (toluene), while other complexes such as **1-Zn** ⊂ C₆₀, in contrast, dissociated completely under the same chromatographic conditions.

For the interaction of **1-RhMe** with fullerenes, the axial methyl groups of the host serve as an NMR probe: the ¹H NMR spectrum of **1-RhMe** alone in C₆D₆ at 25 °C showed eight inequivalent doublets assigned to MeRh at $\delta = -5.15$, -5.30 , -5.36 , -5.67 , -5.77 , -5.79 , -5.84 , and -5.85 , which indicate the presence of several conformational isomers.^[4] When C₆₀ was added to this solution, **1-RhMe** underwent a conformational change to give only a single doublet for MeRh (**1-RhMe** ⊂ C₆₀) at $\delta = -5.06$. In the ¹³C NMR spectrum in CDCl₃, **1-RhMe** ⊂ C₆₀ showed a signal assigned to MeRh in a region downfield (d, $\delta = -6.9$, $\Delta\delta = 6.9$) from that of **1-RhMe** ($\delta = -13.8$), while included C₆₀ displayed a signal in a region upfield ($\delta = 139.4$; $\Delta = -3.6$) from that of free C₆₀ ($\delta = 143.0$). In contrast, upon coordination of an electron-donating ligand such as bipyridine (bpy) to **1-RhMe** (**1-RhMe** ⊂ bpy), the ¹³C NMR signal for MeRh shifted upfield^[14] to $\delta = -16.0$ ($\Delta\delta = -2.2$). Therefore, it is likely that **1-RhMe** ⊂ C₆₀ involves a rhodium-to-fullerene charge transfer interaction.

The much higher affinities of the host molecules toward C₇₀ than C₆₀ are considered to be a result of the ovality of C₇₀ (7.1/7.9 Å). Because of this elliptical shape, C₇₀ itself shows five inequivalent ¹³C NMR signals.^[15, 16] When ¹³C-enriched C₇₀ was mixed with **1-Zn** in CDCl₃/CS₂ (1:1), all the C₇₀ signals shifted upfield, whereby the change in chemical shift was more pronounced for the equatorial carbon atoms than for those at the pole over a wide temperature range ($\Delta\delta_{\text{equator}} = -3.8$, $\Delta\delta_{\text{pole}} = -2.4$ at 25 °C; $\Delta\delta_{\text{equator}} = -4.1$, $\Delta\delta_{\text{pole}} = -2.5$ at -60 °C).^[5, 17] Since the upfield shifts of the fullerene signals are induced mostly by the shielding effect of the porphyrin macrocycle, this trend indicates that included C₇₀ adopts a side-on conformation with respect to the two zinc porphyrins, which possibly maximizes the host–guest π interaction.^[5, 7b] With **1-RhMe** as host, C₇₀ upon inclusion showed a similar change in spectral profile at 25 °C ($\Delta\delta_{\text{equator}} = -3.6$, $\Delta\delta_{\text{pole}} = -3.0$).^[18] However, on lowering the temperature to -60 °C, the shielding effect became slightly but definitely more pronounced for the carbon atoms at the pole than for those on the equator ($\Delta\delta_{\text{equator}} = -3.9$, $\Delta\delta_{\text{pole}} = -4.1$), which suggests that an end-on orientation is preferred for included C₇₀ at low temperatures.^[19]

Variable-temperature (VT) ¹³C NMR spectroscopy is informative of the dynamics of the complexation of fullerenes with cyclic dimers of metalloporphyrins.^[5] For example, a 1:2 mixture of **1-Zn** and ¹³C-enriched C₆₀ at -60 °C in [D₈]toluene showed two singlet ¹³C NMR signals at $\delta = 139.7$ and 142.9 arising from included and free C₆₀ molecules, respectively (Figure 3a). On elevation of the temperature, the signals coalesced at 20 °C as a result of exchange of C₆₀ faster than the NMR timescale. When ¹³C-enriched C₇₀ was the guest ([**1-Zn**]/[C₇₀] = 1:2), a similar coalescence signature was observed at a higher temperature (60 °C), as the K_{assoc} value for C₇₀ is 31 times larger than that for C₆₀. In sharp contrast, although the K_{assoc} value is almost comparable to that for the **1-Zn**/C₇₀

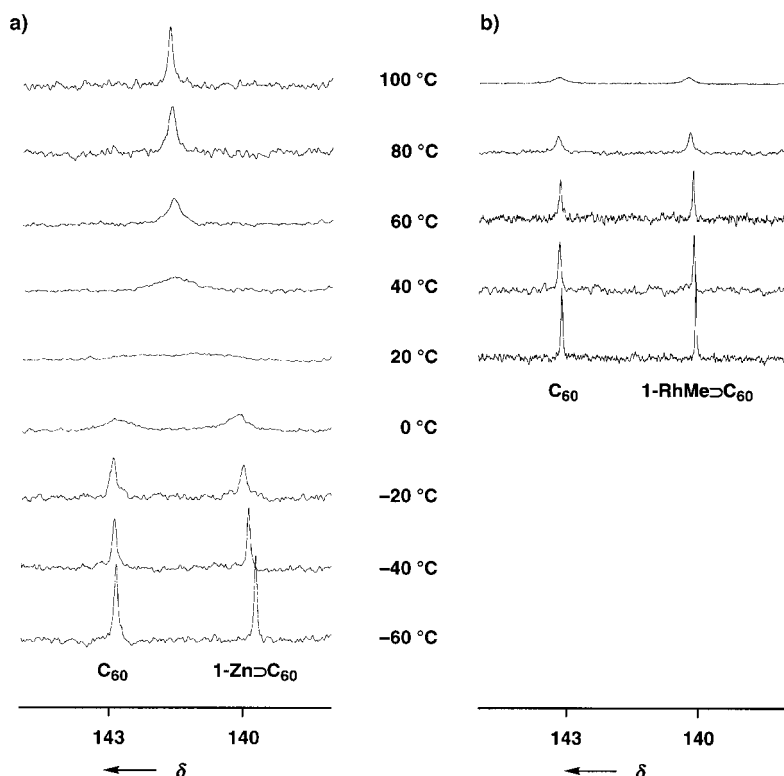


Figure 3. VT ^{13}C NMR profiles of ^{13}C -enriched C_{60} in the presence of 0.5 equiv of **1-Zn** (a) and **1-RhMe** (b) in $[\text{D}_8]\text{toluene}$.

system (Figure 2), the **1-RhMe**/ C_{60} (1:2) system did not show such a coalescence phenomenon even at 100 °C (Figure 3b). The same was true of the **1-RhMe**/ C_{70} system. Since the rate constant of dissociation k_{dissoc} is much smaller than that of association k_{assoc} ($K_{\text{assoc}} = k_{\text{assoc}}/k_{\text{dissoc}} \gg 1$), the above coalescence profiles should reflect the magnitude of k_{dissoc} rather than k_{assoc} . From the differences in resonance frequency between the free and included fullerenes,^[20] the k_{dissoc} values for **1-Zn** \supset C_{60} and **1-Zn** \supset C_{70} at the coalescence temperatures were estimated to be 450 (20 °C) and 310 s^{-1} ^[21] (60 °C), respectively.^[22] The absence of coalescence for the **1-RhMe** fullerene systems indicates that **1-RhMe** \supset fullerenes dissociate much less frequently than **1-Zn** \supset fullerenes. Together with the fact that rhodium(III) porphyrins can form ethene π complexes,^[23] the very low dissociation activities of **1-RhMe** \supset fullerenes suggest some sort of bonding interaction between the central rhodium(III) ions of the host and the included fullerenes.

By taking advantage of the tunable affinity of the host molecule toward fullerenes, choice of appropriate central metal ions of the host allows simple separation of C_{70} from a mixture of C_{60} and C_{70} . For example, a $\text{C}_{60}/\text{C}_{70}$ mixture containing 65 % of C_{70} was mixed with **2-Zn** ([fullerenes]/[**2-Zn**] = 10) in benzene,^[24] and the inclusion complexes were isolated by column chromatography on alumina, and then treated with 4,4'-bipyridine to extract the included fullerenes from the cavity. According to HPLC analysis on Buckyprep with toluene as eluent, the extract contained almost pure C_{70} (> 99 %). Even from a $\text{C}_{60}/\text{C}_{70}$ mixture with a much lower C_{70} content such as 20 %, an extract highly enriched in C_{70} (91 %) was again obtained.

In conclusion, we have demonstrated a remarkable effect of the central metal ions on the interaction of metalloporphyrin cyclic dimers with fullerenes. In particular, the extremely high association constants ($\sim 10^8 \text{ M}^{-1}$)^[25] and low dissociation activities of the **1-RhMe**/fullerene systems should allow construction of more complex supramolecular architectures based on fullerenes. Further studies along this line together with an investigation of electronic and magnetic properties of such multiwalled π -electron supramolecular systems are now in progress.

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- higher than those of **1**-Zn ($1.4 \times 10^4 \text{ M}^{-1}$) and **1**-Co ($7.2 \times 10^4 \text{ M}^{-1}$) under the same conditions.
- [14] ^{13}C NMR (CDCl_3 , 25°C) of bipyridine: $\delta = 150.3$ (C2), 121.1 (C3), 145.2 (C4); **1**-RhMe \supset bipyridine: $\delta = 144.2$ (C2), 117.9 (C3), 140.7 (C4).
- [15] ^{13}C NMR ($\text{CDCl}_3/\text{CS}_2$ 1:1): C_{70} (equator to pole) at 25°C : $\delta = 130.4$, 144.9, 147.6, 146.9, 150.1; at -60°C : $\delta = 130.0$, 144.5, 147.2, 146.5, 149.7; $\text{C}_{70}/\textbf{1}$ -Zn (1:2) at 25°C : $\delta = 126.6$ ($\Delta\delta = -3.8$), 141.3 (-3.6), 144.5 (-3.1), 144.2 (-2.7), 147.7 (-2.4); at -60°C : $\delta = 125.9$ ($\Delta\delta = -4.1$), 140.7 (-3.8), 143.9 (-3.3), 143.7 (-2.8), 147.2 (-2.5).
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Photoelectrochemistry with Controlled DNA-Cross-Linked CdS Nanoparticle Arrays**

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Electronics based on DNA has been the subject of extensive recent research activities that address the conductivity features of double-stranded (ds) DNA,^[1–7] the use of ds-DNA as a template for the construction of nanowires,^[8] and the employment of metal nanoparticles cross-linked by DNA

as single-electron charging devices.^[9, 10] The optical properties of DNA-cross-linked Au nanoparticles were recently studied and applied for DNA sensing,^[11, 12] and nanoarchitectures of DNA/Au nanoparticles were assembled.^[13] The electronic transduction of DNA sensing and, in particular, amplified DNA analyses were recently studied by using electrochemical measurements^[14–23] or microgravimetry^[16] with a quartz-crystal microbalance. Direct electrochemical detection of DNA was achieved by monitoring the electrochemical response of DNA^[14–16] or by the incorporation of redox-labels into ds-DNA.^[17–19] The amplified detection of DNA was accomplished by the use of biocatalytic conjugates^[20, 21] and the application of labeled liposomes^[22, 23] or nanoparticles.^[24, 25]

Herein we describe the novel architecture of double-stranded DNA-cross-linked CdS nanoparticle arrays on electrode supports and the structurally controlled generation of photocurrents upon irradiation of these arrays. The electrostatic binding of $[\text{Ru}(\text{NH}_3)_6]^{3+}$ to the ds-DNA units provides tunneling routes for the conduction-band electrons and thus results in enhanced photocurrents. Besides the unique and novel photoelectrochemical features of the systems, the assemblies provide a new means for the optical fluorescence and electronic (photoelectrochemical) transduction of DNA-sensing events.

CdS nanoparticles ($2.6 \pm 0.4 \text{ nm}$) were functionalized with thiolated oligonucleotide **1** or **2**. These two oligonucleotides are complementary to the 5'- and 3'-ends of the target DNA **3**. The size of the CdS nanoparticles was determined from HR-transmission electron microscopy (TEM) images (average of 150 particles). Using the absorption spectra of the CdS nanoparticles ($\lambda_{\text{max}} = 405 \text{ nm}$) and the Brus equation^[26] gave a similar particle size. We estimate that about 20–24 oligonucleotide units are associated with each CdS nanoparticle (see *Experimental Section* for details). Figure 1 shows the stepwise assembly of the DNA-cross-linked CdS particles on an Au electrode. The oligonucleotide **1** was assembled on the Au electrode ($2.3 \times 10^{-11} \text{ mol cm}^{-2}$) and then treated with the analyte **3** to yield the ds system. Subsequent interaction of the surface with the **2**-functionalized CdS resulted in the binding of the CdS nanoparticles to the surface. Further alternating interaction of the interface with a solution containing the **1**-functionalized CdS nanoparticles (1 mg mL^{-1}) pretreated with **3** ($1 \times 10^{-6} \text{ M}$) and with a solution containing **2**-functionalized CdS nanoparticles resulted in an array with a controlled number of CdS nanoparticle generations. The buildup of the DNA-cross-linked CdS nanoparticle array was monitored by microgravimetry with a quartz-crystal microbalance (Figure 2A).

From the frequency changes observed upon the buildup of the first generation of CdS nanoparticles, and knowing their dimensions, we estimate the surface coverage to be about 1.6×10^{12} particles per square centimeter. This value corresponds to around 15% of a densely packed particle layer. The negative charge associated with the nucleic acid functionalized nanoparticles probably prevents the formation of a dense particle configuration. The DNA-cross-linked CdS nanoparticle arrays were also assembled on glass supports by using an aminopropylsiloxane-functionalized glass that was treated with ϵ -maleimidocaproic *N*-hydroxysuccinimide ester^[24, 25] as a base interface for the covalent linkage of **1** and the organization of the nanoparticle systems. Figure 2B shows

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